

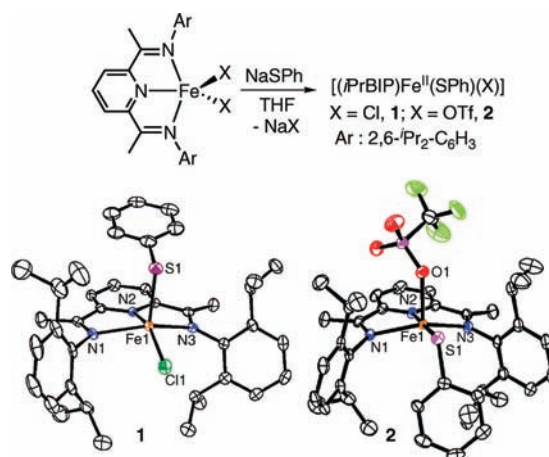
## O<sub>2</sub> Activation by Bis(imino)pyridine Iron(II)–Thiolate Complexes

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**S** Supporting Information

**ABSTRACT:** The new iron(II)–thiolate complexes [(<sup>i</sup>PrBIP)Fe<sup>II</sup>(SPh)(Cl)] (1) and [(<sup>i</sup>PrBIP)Fe<sup>II</sup>(SPh)(OTf)] (2) [BIP = bis(imino)pyridine] were prepared as models for cysteine dioxygenase (CDO), which converts Cys to Cys-SO<sub>2</sub>H at a (His)<sub>3</sub>Fe<sup>II</sup> center. Reaction of 1 and 2 with O<sub>2</sub> leads to Fe-oxygenation and S-oxygenation, respectively. For 1 + O<sub>2</sub>, the spectroscopic and reactivity data, including <sup>18</sup>O isotope studies, are consistent with an assignment of an iron(IV)–oxo complex, [(<sup>i</sup>PrBIP)Fe<sup>IV</sup>(O)(Cl)]<sup>+</sup> (3), as the product of oxygenation. In contrast, 2 + O<sub>2</sub> results in direct S-oxygenation to give a sulfonato product, PhSO<sub>3</sub><sup>-</sup>. The positioning of the thiolate ligand in 1 versus 2 appears to play a critical role in determining the outcome of O<sub>2</sub> activation. The thiolate ligands in 1 and 2 are essential for O<sub>2</sub> reactivity and exhibit an important influence over the Fe<sup>III</sup>/Fe<sup>II</sup> redox potential.



**Figure 1.** Synthetic scheme and displacement ellipsoid plots (50% probability level) for 1 and 2 at 110 K. H atoms have been omitted for clarity.

Determining the factors that govern the activation of dioxygen by both heme and non-heme iron metalloenzymes is of fundamental importance. Mononuclear non-heme iron oxygenases typically contain a 2-His-1-carboxylate ligand set bound to the catalytic iron center. An interesting exception is cysteine dioxygenase (CDO), which utilizes a (His)<sub>3</sub>Fe<sup>II</sup>(H<sub>2</sub>O) center to activate O<sub>2</sub> and oxidize cysteine to sulfenic acid (CysSO<sub>2</sub>H), a key metabolic process that is vital for human health.<sup>1</sup> Despite the importance of CDO from a health perspective, little is known about the mechanism of this dioxygenase.<sup>2</sup> The oxidation of Cys to disulfide, sulfenic acid [Cys(O)H], and other oxidized products has been implicated in oxidative stress response.<sup>3</sup> Thus, understanding the fundamental mechanistic pathways of biologically relevant sulfur oxidations is of high current interest.<sup>4</sup>

Although many studies on iron(II) model complexes have yielded key insights into the reactivity of non-heme iron centers, relatively few have involved the use of O<sub>2</sub> as the oxidant, in part because of the inherent difficulties with activating and controlling O<sub>2</sub>.<sup>5</sup> In an earlier report, we described the synthesis of a N<sub>3</sub>S(thiolate)Fe<sup>II</sup> model complex of CDO that contains the three-neutral-N binding motif found in the enzyme and reacts with O<sub>2</sub> selectively to yield an S-oxygenated sulfonato product.<sup>6</sup> The thiolate donor was covalently tethered to a bis(imino)pyridine (BIP) framework, in part to favor S-oxygenation as opposed to disulfide formation. To our knowledge, this reaction was the first example of an Fe<sup>II</sup>–thiolate complex reacting with O<sub>2</sub> to give S- as opposed to Fe-oxygenation (e.g., Fe<sup>III</sup>–O–Fe<sup>III</sup> species).<sup>7</sup>

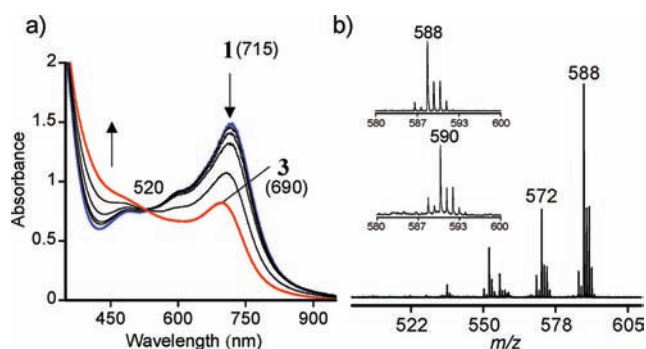
Herein we report the synthesis of two new unsymmetrical Fe<sup>II</sup>–thiolate BIP complexes, [(<sup>i</sup>PrBIP)Fe<sup>II</sup>(SPh)(Cl)] (1) and [(<sup>i</sup>PrBIP)Fe<sup>II</sup>(SPh)(OTf)] (2) [<sup>i</sup>PrBIP = 2,6-(ArN=CMe)<sub>2</sub>-C<sub>5</sub>H<sub>3</sub>N, Ar = 2,6-<sup>i</sup>Pr<sub>2</sub>C<sub>6</sub>H<sub>3</sub>], in which the thiolate ligands are

not covalently tethered to the BIP framework. The reactivity of these complexes toward O<sub>2</sub> has been examined together with non-thiolate-ligated analogues. We show that coordination of the thiolate ligands is crucial for O<sub>2</sub> activation by (BIP)Fe<sup>II</sup>. We also show that S-oxygenation is possible for a terminal thiolate and furthermore that the positioning of the thiolate donor specifies the outcome of oxygenation at either sulfur or iron.

Significant efforts have gone into the synthesis and study of (BIP)Fe complexes for their use in N<sub>2</sub> activation and catalysis.<sup>8</sup> However, unsymmetrical derivatives having the formula [(BIP)Fe<sup>II</sup>(X)(Y)] (X ≠ Y) are scarce. Careful control of stoichiometry, together with the appropriate conditions (solvent, temperature), allowed for the isolation of the monothiolato complexes 1 and 2 (Figure 1). The molecular structures of 1 and 2 reveal five-coordinate Fe<sup>II</sup> ions with the desired single terminal thiolate ligands bound to the iron. The bond distances and angles are consistent with those of high-spin Fe<sup>II</sup> BIP complexes.<sup>8a,d,e</sup> A distinguishing feature of the structures of 1 and 2 is the positioning of the thiolate ligand. In complex 1, the PhS<sup>-</sup> group sits in a pseudoaxial position in relation to the N<sub>3</sub>Cl plane and is oriented trans to the open coordination site that subtends the obtuse N1–Fe–N3 angle (141.2°). This positioning may be aided by a π-stacking interaction between the pyridine and PhS<sup>-</sup> groups. In contrast, the PhS<sup>-</sup> ligand in 2 is bound in a pseudoequatorial arrangement with the <sup>i</sup>PrBIP ligand and is cis to the open coordination site.

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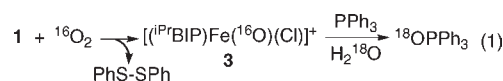
**Figure 2.** (a) UV-vis spectral changes for the reaction of **1** (715 nm, 0.37 mM) with excess  $O_2$  in  $CH_2Cl_2$ , leading to formation of **3** (690 nm). (b) LDIMS of **3** formed in the reaction of **1** +  $O_2$ . Peaks at  $m/z$  588 and 572 correspond to  $[(^{iPr}BIP)Fe^{IV}(O)(Cl)]^+$  and  $[(^{iPr}BIP)Fe^{II}(Cl)]^+$ , respectively. Inset: isotopic clusters of **3** prepared from  $^{16}O_2$  (top) and  $^{18}O_2$  (bottom).

Both **1** and **2** exhibit relatively sharp, paramagnetically shifted peaks in the  $^1H$  NMR spectrum ( $CD_2Cl_2$ ) typical of high-spin (BIP)FeX<sub>2</sub> complexes, and these spectra are consistent with their solid-state structures. The magnetic susceptibility of **1** measured by Evan's method in  $CD_2Cl_2$  gave  $\mu_{eff} = 5.2 \mu_B$ , which is close to the spin-only value for a high-spin  $Fe^{II}$  ( $S = 2$ ) ion.

Reaction of **1** with a slight excess of dry  $O_2$  (5 equiv) led to a color change from dark-blue to green over the course of 1 h. A decrease in the band at  $\lambda_{max} = 715$  nm for **1** ( $\epsilon \approx 4000 M^{-1} cm^{-1}$ ) was observed, and a new band for the green species appeared at  $\lambda_{max} = 690$  nm ( $\epsilon \approx 1500 M^{-1} cm^{-1}$ ) (Figure 2; for the time dependence, see Figure S5 in the Supporting Information). This spectrum is similar to that reported for a closely related bis(imino)pyridine iron(IV)-oxo complex ( $\lambda_{max} 660$  nm,  $\epsilon \approx 1200 M^{-1} cm^{-1}$ ).<sup>9</sup> Analysis by laser desorption ionization mass spectrometry [LDIMS(+)] revealed a dominant isotopic cluster at  $m/z$  588 whose isotope and fragmentation pattern (Figure 2 and Figures S8 and S9) are consistent with an  $Fe^{IV}(O)$  complex,  $[(^{iPr}BIP)Fe^{IV}(O)(Cl)]^+$  (**3**). The thiolate ligand is oxidized to disulfide during the production of **3**, as determined by  $^1H$  NMR spectroscopy (PhS-SPh, 85%). Introduction of  $^{18}O_2$  in place of  $^{16}O_2$  caused a shift of two mass units for the LDIMS of **3**, giving a peak at  $m/z$  590 (80%  $^{18}O$  incorporation). Finally, green **3** was EPR-silent (X-band, 15 K). These data are consistent with the assignment of **3** as an  $Fe^{IV}(O)$  species.

When the reaction of **1** with excess  $O_2$  in  $CH_2Cl_2$  was carried out in the presence of  $PPh_3$  (5 equiv),  $OPPh_3$  was produced in good yield (70% by  $^{31}P$  NMR analysis). Alternatively, formation of green **3** followed by removal of  $O_2$  under vacuum and addition of  $PPh_3$  (50–300 equiv) under Ar resulted in the smooth decay of the peak for **3** at 690 nm (Figure S6). This decay followed good pseudo-first-order behavior, and the rate constants ( $k_{obs}$ ) thus obtained were found to increase linearly with  $[PPh_3]$ , yielding a second-order rate constant of  $k_2 = (3.6 \pm 0.3) \times 10^{-3} M^{-1} s^{-1}$  for oxygen atom transfer from **3** to  $PPh_3$  (Figure S7). This relatively slow reactivity may be due to the steric encumbrance imposed by the 2,6- $iPr_2C_6H_3$  substituents. The  $^{18}O$ -labeled **3** produced  $^{18}OPPh_3$  with modest isotopic incorporation ( $^{16}O/^{18}O = 85:15$ ). However, addition of excess  $H_2^{18}O$  to the reaction of **3**- $^{16}O$  and

$PPh_3$  resulted in a significant increase in the isotopically labeled product  $^{18}OPPh_3$  (50%  $^{18}O$ ) (eq 1):

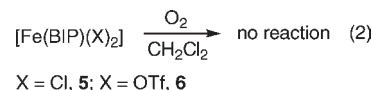


These data indicate that the O atom in **3** undergoes facile exchange with exogenous  $H_2O$ , as seen for other terminal iron-oxo species.<sup>10</sup> Although further spectroscopic studies are needed to definitively characterize the structure of **3**, all of the spectroscopic data and reactivity presented here strongly support the formulation of **3** as a terminal iron-oxo complex generated from **1** +  $O_2$ , with the PhS<sup>−</sup> ligand undergoing concomitant oxidation to disulfide.

The formation of non-heme  $Fe^{IV}(O)$  complexes from  $Fe^{II}$  and  $O_2$  can be induced by the addition of external coreductants (e.g., cyclohexene or NADH).<sup>5b,c,e</sup> In the case of **1**, the thiolate ligand functions as a built-in coreductant to assist in the activation of  $O_2$ . In comparison, the covalently tethered thiolate complex  $[Fe^{II}(N_3S(thiolate))(OTf)]$  (**4**) also serves to activate dioxygen, but in that case, participation from sulfur leads to direct oxygenation of the S atom.<sup>6</sup>

To our surprise, the addition of stoichiometric amounts of  $O_2$  to the triflate complex **2** followed a very different oxidation pathway than the one followed by the chloro analogue **1**. An immediate color change from dark-blue to brown was noted upon addition of  $O_2$ , and LDIMS revealed a cluster at  $m/z$  694 corresponding to *S*-oxygenated  $[Fe^{II}(^{iPr}BIP)(PhSO_3)]^+$ . Attempts to crystallize  $[Fe^{II}(BIP)(PhSO_3)]^+$  to date have led only to the crystallization of the known  $Fe^{II}(^{iPr}BIP)(OTf)_2$  complex; however, the production of benzenesulfonic acid was readily confirmed by  $^1H$  NMR spectroscopy, and quantitation by reversed-phase HPLC after hydrolytic workup gave a yield of 30% for  $PhSO_3H$  (based on total Fe). The use of labeled  $^{18}O_2$  resulted in  $\sim 90\%$  incorporation of  $^{18}O$  into the  $PhSO_3^-$  ligand. Despite the fact that the thiolate donor in **2** is not part of a chelate ring, *S*-oxygenation does occur, as seen for the covalently tethered **4**. In contrast, no evidence for  $PhSO_3H$  was detected by LDIMS or HPLC for **1** +  $O_2$  in control experiments.

The reactivities of the related non-thiolate-ligated complexes  $Fe^{II}(^{iPr}BIP)Cl_2$  (**5**) and  $Fe^{II}(^{iPr}BIP)(OTf)_2$  (**6**) were next examined for comparison with **1** and **2**. These complexes are completely inert toward  $O_2$  in both solution (e.g.,  $CH_2Cl_2$ ,  $CH_3CN$ ) and the solid state (eq 2):



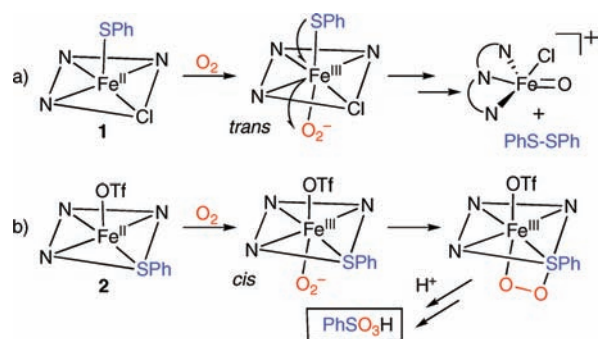
Addition of  $PPh_3$  to oxygenated solutions of **5** and **6** showed no formation of  $OPPh_3$ . The incorporation of a thiolate donor thus clearly plays a critical role in the activation of  $O_2$  by these non-heme iron(II) complexes.

The redox potentials of **1**, **2**, **5**, and **6** are compared in Table 1. The thiolate-ligated complexes exhibit significantly lower redox potentials than the nonthiolate analogues, correlating nicely with their relative  $O_2$  reactivities. A similar correlation was made for  $[Fe^{II}(TMC)(OTf)_2]$  (TMC = 1,4,8,11-tetramethyl-1,4,8,11-tetraazacyclotetradecane), which exhibits a solvent-dependent redox potential and reacts with  $O_2$  to give an  $Fe^{IV}(O)$  complex only in solvents where  $E_{1/2}(Fe^{III/II}) < -0.1$  V (e.g., THF).<sup>5a</sup> Similarly, non-heme iron(II) complexes with more positive  $E_{1/2}$  values fail to react with  $O_2$  to give oxoiron(IV) species. An  $E_{1/2}(Fe^{III/II}) < -0.1$  V appears to be a prerequisite for  $O_2$  activation in non-heme

**Table 1. Redox Potentials for (<sup>iPr</sup>BIP)Fe<sup>II</sup> and Related Non-Heme Fe<sup>II</sup> Complexes**

compound	$E_{1/2}$ ( $\Delta E_{pp}$ ) <sup>a</sup>	O <sub>2</sub> reactivity
[( <sup>iPr</sup> BIP)Fe <sup>II</sup> (SPh)(Cl)] (1)	-0.173 <sup>b</sup> (0.114) (r)	yes
[( <sup>iPr</sup> BIP)Fe <sup>II</sup> (SPh)(OTf)] (2)	-0.372 <sup>b</sup> (0.149) (r)	yes
[( <sup>iPr</sup> BIP)Fe <sup>II</sup> (Cl) <sub>2</sub> ] (5)	0.025 <sup>b</sup> (0.153) (r)	no
[( <sup>iPr</sup> BIP)Fe <sup>II</sup> (OTf) <sub>2</sub> ] (6)	0.613 <sup>b,c</sup> (ir)	no
[(TMC)Fe <sup>II</sup> (OTf) <sub>2</sub> ] <sup>d</sup>	-0.14 <sup>e</sup> (qr)	yes
[(TMC)Fe <sup>II</sup> (OTf) <sub>2</sub> ] <sup>d</sup>	0.02 <sup>f</sup> (r)	no
[(TPA)Fe <sup>II</sup> ] <sup>2+</sup> , <sup>d,g</sup>	0.36 <sup>h</sup> (r)	no

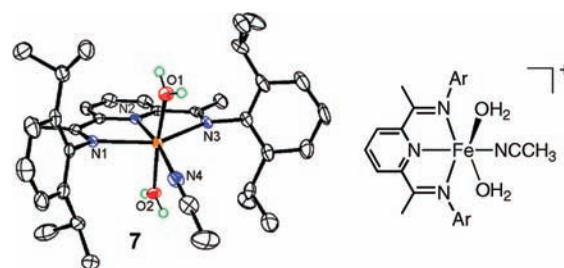
<sup>a</sup> Values in V vs Fc<sup>+</sup>/Fc.  $\Delta E_{pp}$  = peak-to-peak separation; r = reversible, ir = irreversible, qr = quasi-reversible. <sup>b</sup> In CH<sub>2</sub>Cl<sub>2</sub> at a scan rate of 0.1 V/s. <sup>c</sup> Anodic peak potential. <sup>d</sup> Data taken from ref 5a. <sup>e</sup> In 1:1 MeCN/THF. <sup>f</sup> In 1:1 MeCN/CH<sub>2</sub>Cl<sub>2</sub>. <sup>g</sup> TPA = tris(2-pyridylmethyl)amine. <sup>h</sup> In neat MeCN.

**Scheme 1. Proposed Mechanisms of O<sub>2</sub> Activation by 1 and 2**

iron(II) complexes, and inclusion of a single thiolate donor is sufficient to lower the redox potential of (<sup>iPr</sup>BIP)Fe<sup>II</sup> complexes into this range. It should be noted that the  $E_{1/2}$  values for **1** and **2** remain more than 1 V above the one-electron reduction potential for the O<sub>2</sub>/O<sub>2</sub><sup>-</sup> couple in organic solvents,<sup>5h</sup> ruling out an outer-sphere mechanism for O<sub>2</sub> activation.

In view of the structural and electronic similarities between the two thiolate-ligated complexes **1** and **2**, why do their reactivities with O<sub>2</sub> follow such dramatically different paths? Scrutiny of the structures of **1** and **2** appears to hold the key. The PhS<sup>-</sup> ligand in **1** is bound trans to the open site available for O<sub>2</sub> binding, whereas it is bound cis in **2**. A plausible mechanism for O<sub>2</sub> activation in **1** thus begins with coordination of O<sub>2</sub> to the open site trans to the thiolate donor, which is followed by electron transfer from both the iron and sulfur centers to the bound O<sub>2</sub> (Scheme 1a). In this case, intramolecular attack of an Fe–O<sub>2</sub> intermediate on the sulfur donor would be strongly disfavored by the trans orientation of the PhS<sup>-</sup> ligand. In contrast, the analogous Fe–O<sub>2</sub> intermediate in **2** would be generated cis to the thiolate ligand, providing a facile pathway for intramolecular S-oxygenation (Scheme 1b). Similarly, the thiolate donor in the covalently tethered **4** is also found cis to the open coordination site.

This hypothesis depends upon the feasibility of attaining a six-coordinate structure with the sterically encumbered BIP ligand in **1** and **2**. For less bulky BIP analogues, where Ar = 2,6-Me<sub>2</sub>C<sub>6</sub>H<sub>3</sub>, six-coordinate Fe<sup>II</sup> complexes are known,<sup>8d</sup> but to our knowledge there are no examples with Ar = 2,6-<sup>iPr</sup>Pr<sub>2</sub>C<sub>6</sub>H<sub>3</sub>. Thus we were pleased to isolate [Fe<sup>II</sup>(<sup>iPr</sup>BIP)(H<sub>2</sub>O)<sub>2</sub>(NCCH<sub>3</sub>)](OTf)<sub>2</sub> (**7**) as a product from the reaction of **2** + O<sub>2</sub>; its molecular structure is given in



**Figure 3.** (left) Displacement ellipsoid plot (50% probability level) at 110 K and (right) molecular structure of **7**. H atoms (except those attached to the water molecules) and the OTf<sup>-</sup> ions have been omitted for clarity.

**Figure 3.** Despite the large steric encumbrance provided by the flanking 2,6-<sup>iPr</sup>Pr<sub>2</sub>C<sub>6</sub>H<sub>3</sub> substituents, a six-coordinate geometry is clearly attainable in **7**.

In summary, we have demonstrated that a thiolate donor is essential for the activation of O<sub>2</sub> by non-heme iron (BIP)Fe<sup>II</sup> complexes and can serve as either a coreductant or a site for O capture. The relative positioning of the PhS<sup>-</sup> ligand in relation to the potential O<sub>2</sub> binding site appears to play a critical role in determining whether oxygenation occurs at iron or sulfur.<sup>11</sup> We have also shown that S-oxygenation can occur for terminal, iron-bound thiolates, contrary to established precedent. It has been proposed that the Cys substrate in CDO coordinates to the Fe center through a chelate ring involving sulfur and the amino group.<sup>1</sup> The findings presented here suggest that this unusual binding mode for Cys is not required for S-oxygenation to occur.

**ASSOCIATED CONTENT**

**S Supporting Information.** Experimental procedures and characterization data for **1–3**, **7** and X-ray crystallography details and CIFs for **1**, **2**, and **7**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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